Corneal cross-linking combines short wavelength light and a chromophore to induce specific changes in the collagen of the corneal stroma. The first change that was investigated in detail was the increase in the biomechanical stability of the cornea, and this specific property of CXL was used to develop therapeutic strategies for corneal ectasias, like keratoconus and postoperative ectasia. Another CXL-mediated effect includes changes in the tertiary structure that affect corneal packing density and swelling behavior. This property was used to develop treatment modalities for bullous keratopathy, early Fuch’s dystrophy, and corneal edema. Finally, CXL increases the corneal resistance to enzymatic digestion by steric hindrance, a mechanism used to treat infectious and noninfectious corneal melting.

In certain conditions, CXL must be applied very close to the corneal limbus. Such conditions include pellucid marginal degeneration (PMD), Terrien’s marginal degeneration, and also rosacea-mediated or trophic peripheral ulcers. Here, the UV-A irradiation must be performed in a decentered manner to the periphery of the lower cornea, partly irradiating the corneal limbus. Here, we investigated the effect of standard and double-standard fluence corneal cross-linking with riboflavin and UV-A (CXL) on cornea and corneal limbus in the rabbit eye in vivo.

METHODS. Epithelium-off CXL was performed in male New Zealand White rabbits with two irradiation diameters (7 mm central cornea, 13 mm cornea and limbus), using standard fluence (5.4 J/cm²) and double-standard fluence (10.8 J/cm²) settings. Controls were subjected to epithelial removal and riboflavin instillation, but were not irradiated with UV-A. Following CXL, animals were examined daily until complete closure of the epithelium, and at 7, 14, 21, and 28 days. Animals were killed and a corneoscleral button was excised and processed for light microscopy and immunohistochemistry.

RESULTS. For both irradiation diameters and fluences tested, no signs of endothelial damage or limbal vessel thrombosis were observed, and time to re-epithelialization was similar to untreated controls. Histological and immunohistochemical analysis revealed no differences in the p63 putative stem cell marker expression pattern.

CONCLUSIONS. Even when using fluence twice as high as the one used in current clinical CXL settings, circumferential UV-A irradiation of the corneal limbus does not alter the regenerative capacity of the limbal epithelial cells, and the expression pattern of the putative stem cell marker p63 remains unchanged. This suggests that eccentric CXL may be performed safely in PMD.

Keywords: corneal cross-linking, high fluence, limbal stem cells, epithelial regeneration, p63...
and proparacaine eye drops were applied topically. CXL was performed as described previously. Briefly, corneas were de-epithelialized mechanically in a diameter of 10 to 11 mm, leaving a margin of 1 mm to the limbus. Isotonic 0.1% riboflavin solution containing 20% dextrane (MedioCross D Solution; Peschke Meditrade GmbH, Hünenberg, Switzerland) was instilled every 2 minutes for 25 minutes without the use of a speculum to avoid corneal dehydration and stromal thinning. Corneal thickness was verified immediately before irradiation by ultrasound pachymetry (Tomey SP-2000; Tomey Corporation), and all corneas used in the experiments showed no more than ±10% variation of central thickness when compared with thickness before riboflavin instillation. The Table shows experimental irradiation and fluence settings: UV-A irradiation was performed in the right eye at 365 nm with a fluence of either 5.4 J/cm² (5 mW/cm² for 30 minutes) or 10.8 J/cm² (9 mW/cm² for 20 minutes) using a commercially available device (CXL-365; Schwind Eye-Tech-Solutions, Kleinostheim, Germany). The irradiation zone was either 7 mm centered on the cornea, or 13 mm, including the entire cornea and limbal area. Following CXL, ofloxacin ointment was administered three times daily until complete closure of the corneal epithelium. Animals were examined daily and the speed and extent of re-epithelialization was documented. The left eyes served as controls. Controls were treated similarly, but not irradiated with UV-A. Animals were killed 4 weeks after CXL and corneoscleral buttons were excised.

### Results

Figure 1 shows the speed of re-epithelialization in corneas irradiated on 13 mm with a fluence of 5.4 J/cm² and 10.8 J/cm², and in unirradiated controls. When using the double-standard fluence of 10.8 J/cm², corneas showed a partial re-epithelialization at 24 hours (Fig. 1B) and a complete re-epithelialization at 72 hours (Fig. 1C) after CXL. Corneas that were treated using the standard fluence of 5.4 J/cm² (data not shown) and the unirradiated controls (Figs. 1E, 1F) showed a re-epithelialization speed at 24 hours and 72 hours, similar to the one observed at 10.8 J/cm² (double-standard fluence).

Light microscopic analysis of the cornea and the limbal area was performed at 4 weeks after CXL. Hematoxylin and eosin staining showed a regular morphology of the corneal stroma as well as of the corneal and limbal epithelial cells after irradiation at a fluence of 5.4 J/cm² (Figs. 2C, 2D) and 10.8 J/cm² (Figs. 2E, 2F).

Immunohistochemical analysis of the corneal-specific marker cytokeratin 3 (K3) showed similar expression in all layers of the corneal epithelium and in the suprabasal limbal epithelium for all conditions tested (Figs. 2F–K).

### Discussion

Currently, the most common indication for CXL is classic progressive keratoconus with stromal thinning and bulging, usually occurring in the midperiphery of the cornea, 3 to 4 mm from the center. Accordingly, irradiation is performed in a centered fashion over an area of 8 to 9 mm. Besides keratoconus, CXL was successfully applied in a number of other corneal diseases like PMD, Terrien’s marginal degeneration, and rosacea-mediated or trophic peripheral ulcers. All conditions have in common that they affect the periphery of the cornea rather than the center or the midperiphery. Here, irradiation must be decentralized and applied in an eccentric fashion. Partial irradiation of the limbus is unavoidable, even when attempts are made to shield the limbus.

Concerning PMD, only a few cases treated by CXL have been reported in the literature so far. Steppat and colleagues (Steppat M, et al. IOVS 2008;49:ARVO E-Abstract 4342) reported on 13 eyes of eight patients with PMD and a clinical follow-up of 18 months. They found an arrest of PMD progression in all eyes in the absence of side effects. In particular, no clinical signs of limbal stem cell insufficiency were observed. Spadaccca described a 43-year-old patient with bilateral PMD who underwent successful CXL in his left eye. Hassan and colleagues reported on a similar case of bilateral PMD where one eye was successfully treated with CXL. Kaya et al. reported a case of PMD in a case series of patients with ectasia and thin corneas who was successfully treated with CXL. Finally, Hafezi et al. described a case of effective CXL for postoperative ectasia with underlying forme fruste PMD.

### Table

<table>
<thead>
<tr>
<th>Group</th>
<th>No. of Animals</th>
<th>Fluence (Intensity × Time)</th>
<th>Irradiation Pattern</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>4</td>
<td>5.4 J/cm² (9 mW/cm² for 10 min)</td>
<td>Cornea only (7 mm)</td>
</tr>
<tr>
<td>2</td>
<td>4</td>
<td>10.8 J/cm² (9 mW/cm² for 20 min)</td>
<td>Cornea only (7 mm)</td>
</tr>
<tr>
<td>3</td>
<td>4</td>
<td>5.4 J/cm² (9 mW/cm² for 10 min)</td>
<td>Cornea and limbus (15 mm)</td>
</tr>
<tr>
<td>4</td>
<td>4</td>
<td>10.8 J/cm² (9 mW/cm² for 20 min)</td>
<td>Cornea and limbus (15 mm)</td>
</tr>
</tbody>
</table>

Peroxidase Substrate kit (Vector Laboratories). Sections were washed in tap water, cleared, mounted, and examined on a light microscope (Leica DM1000B; Leica Microsystems, Wetzlar, Germany) using Diskus 4.80 image analyzer software (Leica Microsystems).
In all cases reported, the standard fluence of 5.4 J/cm² was used, at time and energy settings of 30 minutes and 3 mW/cm². Also, all cases reported that re-epithelialization was uneventful and occurred within 4 days, in the absence of side effects. Spadae et al.⁵² and Hassan et al.⁵⁶ reported that they had performed CXL eccentrically by decentering the irradiation zone toward the inferior periphery of the cornea. The question remains as to whether the limbal stem cells tolerate the UV-A irradiation.

**Figure 1.** Re-epithelialization following CXL on 13 mm using standard (5.4 J/cm²) and double-standard fluence (10.8 J/cm²). The erosion was documented immediately after CXL (B, C). At 24 hours (E, F) and 48 hours (H, J) after CXL, re-epithelialization speed was similar for both fluences tested. At 72 hours after CXL, the corneal surface was fully re-epithelialized in standard and double-standard fluence corneas (L, M). The nonirradiated controls showed a similar time to re-epithelialization (A, D, G, K).

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**Figure 2.** Light microscopic and immunohistochemical analysis of cornea and limbus. Corneal histology is regular at a fluence of 5.4 J/cm² when irradiating the cornea at a diameter of 7 mm (B) and cornea and limbus at a diameter of 13 mm (C), and similar to the 7-mm (D) and 13-mm (E) irradiations using double-standard fluence (10.8 J/cm²) and controls (A). Cytokeratin 3 (K3) is predominantly expressed in all layers of the corneal epithelium and in the superficial limbal epithelium and its expression pattern is similar in controls (F), at 5.4 J/cm² using 7-mm (G) and 13-mm (H) irradiation, and at 5.4 J/cm² using 7-mm (J) and 13-mm (K) irradiation. The expression pattern of the stem cell marker p63 is also similar for all conditions tested (L–P).
Other diseases of the peripheral cornea in which CXL was applied include Terrien’s marginal degeneration, and rosacea-mediated or trophic peripheral ulcers. In these cases, a decentered irradiation also was performed to cover the area of active disease. No limbal stem cell deficiency was reported by the authors. The corneal epithelium shows a physiological turnover time of 7 to 14 days and also a distinct renewal capacity. 28,20,31 Cotsarelis and colleagues were the first to identify slow-cycling cells in the mouse limbus that might be responsible for this renewal capacity. A number of specific stem cell markers are known today. The cytokeratin K3 represents a corneal-specific marker that is usually expressed by all corneal epithelia and the suprabasal limbal epithelial cells and was first described by Schermer and colleagues. A marker for limbal epithelial stem cells is the transcription factor p63, which is highly expressed in the basal cell layer, and to a lesser extent in the superficial layers of the limbal epithelial cells. The p63 is not expressed in the corneal epithelium. In our study, we observed an expression pattern similar to untreated controls for all conditions tested (Fig. 2). Accordingly, the speed of epithelial regeneration was similar to controls for both fluences (Fig. 1 and data not shown).

When a sufficient number of stem cells is irreversibly damaged, limbal stem cell deficiency may occur, characterized by a substitution of the corneal epithelium by conjunctival epithelium and leading to corneal opacification. Stem cells may be damaged by several mechanisms, including chemical burns, inflammatory disease, and light. Short-wavelength UV-B (290–320 nm) and UV-A (320–400 nm) light can induce mutations and apoptosis and damage various ocular structures. In the cornea, UV-A at 365 nm is absorbed by the corneal epithelium and can induce direct damage via oxidative stress.

All currently available CXL systems and protocols use a fluence of 5.4 J/cm² and adhere to the Bunson-Roscoe law, varying time and energy settings (30 minutes at 3 mW/cm², 10 minutes at 9 mW/cm², 5 minutes at 18 mW/cm², and 2.5 minutes at 36 mW/cm²). In our study, and in contrast to the studies reported so far, we also applied a fluence of 10.8 J/cm² (20 minutes at 9 mW/cm²). This fluence is twice as high as the one used in all current clinical settings. We deliberately used such a high fluence to test the limits of resistance of the limbal stem cells to irradiation-induced damage. Such a high dose of energy applied to the human eye would inevitably lead to decompensation of the corneal endothelium, since the threshold of endothelial damage of 0.65J/cm² would be exceeded. The rabbit cornea is distinctly thicker than the human cornea and at a thickness of 700 μm, the surface dose at the level of the endothelium is at 0.47 J/cm² only, well below the damage threshold indicated above.

Others also have investigated the effect of CXL on the corneal limbus: in 2011, Wollensak and colleagues performed cell counts and RT-PCR for stem cell markers as well as flow cytometry for a 360-degree UV-A irradiation of the limbus using 5.4 J/cm² (standard CXL) and 10.8 J/cm² (twice the standard fluence currently used in clinical settings), when compared with the epithelial healing rate of conventional CXL with irradiation of the cornea only. Also, CXL has been in clinical use since 1999 and, to date, no cases of limbal stem cell deficiency and/or mutagenesis have been reported. On the other hand, even longer follow-ups might be needed to exclude any long-term mutagenic effects.

In conclusion, we show that the regenerative capacity and the structural integrity of the corneal limbus are preserved after a 360-degree UV-A irradiation of the limbus using 5.4 J/cm² with irradiation of the inferior limbus, which is highly expressed in the basal cell layer, and to a lesser extent in the superficial layers of the limbal epithelial cells. The p63 is not expressed in the corneal epithelium. In our study, we observed an expression pattern similar to untreated controls for all conditions tested (Fig. 2). Accordingly, the speed of epithelial regeneration was similar to controls for both fluences (Fig. 1 and data not shown).

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References


